

International Journal of Botany Studies www.botanyjournals.com

ISSN: 2455-541X

Received: 08-06-2021, Accepted: 23-06-2021, Published: 10-07-2021

Volume 6, Issue 4, 2021, Page No. 134-137

In vitro antimicrobial activities of vegetables (Potato, Cucumber, Sweet Potato and Ginger) peel wastes for ecofriendly microbial management

Raj Singh, Mahiti Gupta, Paavan Singhal, Soniya Goyal, Sushil Kumar Upadhyay*

Department of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Mullana-Ambala, Haryana, India

Abstract

Vegetable processing generates tremendous amount of peel waste, which is dumped in the environment. The vegetables peels are generally considered as waste product and are normally thrown away. These wastes have potential microcidal activities. These can be used as natural and economic source of antimicrobial agent against many pathogenic microbes. In the present study antimicrobial potential of four copiously available vegetable peel wastes (e.g., potato, cucumber, sweet potato and ginger) was evaluated on selected bacterial and fungal strains. The *in vitro* antimicrobial activities were worked out using diffusion assay expressed in terms of zone of inhibition and substantiated by advanced numerical tools. The highest antibacterial activity was found with cucumber against *Escherichia coli* followed by *Staphylococcus aureus*, *Lactobacillus* and *Proteus vulgaris* the highest antifungal activity was found with sweet potato followed by ginger and potato against *Saccharomyces cerevisiae* in terms of zone of inhibition. Hence it confirms the potential of above vegetable peel waste to be used for therapeutic purpose to combat the pathogenic microbes. The studies conducted on peels revealed the presence of important pharmacologically active antimicrobial constituents and suggests future scope of vegetable peel wastes for therapeutic purpose. Thus the ecofriendly approach of microbial management will be helpful in lowering of organic wastes and sustainability of the environment.

Keywords: vegetable peels, *escherichia coli*, *staphylococcus aureus*, *lactobacillus*, *proteus vulgaris*, *saccharomyces cerevisiae*, antimicrobial, microbial management

Introduction

The scientific investigations revealed the uninterrupted rich sources of antimicrobials among vegetables seeds and peels. Usually vegetables skins are thrown in the garbage or fed to livestock. These parts are very rich source of many bioactive components and considered to have antimicrobial potential (Singh et al., 2017) [23]. Therefore, it can be used for pharmacological or pharmaceutical purposes. Various components having antimicrobial, antioxidant and antiinflammatory activities are extracted from different peels (Sehrawat et al., 2020; Yadav et al., 2020) [20, 39]. This prospect will make available an alternative to solve the problems in areas of antibiotic application as an economical and ecofriendly approach in reducing the pollution due to disposal of such agro wastes (Saleem and Saeed, 2020) [18]. It was reported that vegetable wastes of potato, brinial. pumpkin, cauliflower and cabbage, produced proteases during by solid state fermentation used in vitro management of Aspergillus niger (Madhumithah et al., 2011) [11]. The antimicrobial activity of wax gourd (Benincasa hispida) extracts was tested against six Gram positive (Bacillus subtilis, Staphylococcus aureus, Micrococcus species, Staphylococcus epidermidis, Staphylococcus xylosus and Bacillus circulans) and seven Gram-negative bacteria (Salmonella typhimurium, Pseudomonas aeruginosa, Proteus vulgaris, Serratia liquefaciens, Cronobacter muytjens, Shigella boydii and Serratia marcescens), one yeast and two molds using the disc diffusion method and revealed significant antimicrobial potential (Abdullah et al., 2012; Doharey et al., 2021) [1, 9]. In another report the peel extracts of Citrus limon showed maximum antimicrobial activity followed by Manilkara zapota and Carica papaya,

indicating its effectiveness as a promising source of natural antimicrobics (Rakholiya et al., 2014) [17]. There were various fruits (peel, flesh or seed) have been used in traditional medicine for stomachache, sore eyes, fever, diabetes, cardioprotective, hepatoprotective renoprotective purposes (Singh et al., 2018a; Singh et al., 2019; Singh *et al.*, 2020a; Singh *et al.*, 2020b; Singh *et al.*, 2020c) [26, 28, 29, 30, 31]. Papaya has been shown to contain sulphydroxyl protease which can inhibit viralsor microbial infection. These compounds are playing an important role in fruits protection against pathogenic agents responsible for fruit lysis (Rajashekhara et al., 1990) [16]. Phenolic compounds from spices such as zingerone and capsaicin have been found to inhibit the germination of bacterial spores, and many known to be potential anticancer agent as well (Burt, 2004; Tuli et al., 2021) [7, 35].

The wastes from fruits and vegetable processing industries being rich in polysaccharides (cellulose, hemicellulose and lignin) can be subjected to solid state fermentation (SSF) for the production of ethanol which has several uses (Abu Yazid et al., 2017; Mahato et al., 2021) [2, 12]. It can be used as a liquid fuel or liquid fuel supplement and as a solvent in many industries. Citric, succinic, malic, acetic, and tartaric acids are commonly found in fruits and fresh-cut byproducts (Yadav et al., 2021) [40]. They have been traditionally used in the food industry as preservative agents, attributing their antimicrobial efficacy to the pH changes of the treated media. In general, bacteria grow at a pH close to 6.5 to 7.5, but tolerate a pH range from 4 to 9. Yeasts are more tolerant to low pH values than bacteria, whereas molds can grow in the widest pH range. One effective way of limiting microbial growth is increasing the acidity of a particular food by adding an acidic substance (Massilia et al., 2009) [13]. Acids attack cell walls, cell membranes, metabolic enzymes, protein synthesis systems, and the genetic material of microorganisms (Tripathi and Dubey, 2004) [33]. Flavonoids have been reported to enhance the antibacterial, antiviral, or anticancer activities of compounds such as naringenin, acycloguanosine, and tamoxifen (Bracke et al., 1999, Singh et al., 2021a; Tuli et al., 2021) [6, 25, 35]. The mixture of phytochemical constituents in plant extracts can be an advantage due to the synergistic effect that the constituents may have (Bakkali et al., 2008; Singh et al., 2018b; Sheokand et al., 2019; Devi et al., 2020; Singh et al., 2021b) [5, 27, 2, 8, 32]. Present study designed to investigates the antimicrobial activity of potato, cucumber, sweet potato and ginger peel extracts against selected microorganisms. Furthermore, this study also aimed to suggest a solution for agro waste minimization. The results obtained in this study will help to explore the possibility for future use of selected fruit peel wastes as antimicrobial agents.

Materials and Methods

a. Collection and preparation of peel extracts: This study had carried out in the Department of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Mullana-Ambala (Haryana). The vegetable *viz.* potato, cucumber, sweet potato and ginger peel were obtained from local market and some from home waste peel material as well. The vegetables were washed and cleaned very well. Thereafter peeled extraneous matters were dried at room temperature in a shaded region for 7 to 10 days. The dried peels were grinded in mortar and pestle followed by preparation of aqueous extract using sterile distilled water (10 g of each powdered sample in 100 ml of aqueous solvent). The solutions were kept in shaking incubator for 24hrs and filtrate was collected using Whatman No.1 filter

- paper. The collected filtrate was stored at 4°C for further application.
- In vitro antimicrobial activity: Pure culture viz. Escherichia coli, Staphylococcus aureus, Proteus vulgaris, Lactobacillus and Saccharomyces cerevisiae were collected from the Microbiology Laboratory, Department of Biotechnology, MMDU-Mullana. The minimum inhibitory concentration of crude plant extracts against pathogenic strains was determined by dilution method (Sathyabama et al., 2011) [19]. The nutrient agar medium was used for mass culture of microorganisms. Nutrient broth and potato dextrose media was used for the inoculation and standardisation of the microorganisms. The crude plant extract prepared were tested for the antibacterial activity through well diffusion method (Upadhyay, 2016a) [36]. In each well 100µl of crude peel extract was added (Upadhyay, 2016b) [37]. Simultaneously the positive control streptomycin (1mg/ml) applied for gram positive and gram negative bacteria; however, fluconazole (1mg/ml) used against fungi.
- c. Incubation and inhibition analysis: The microbial growth inhibition zone for bacteria was measured after incubation at 37°C. While in case of fungi it was measured after incubation at 30°C. The appearance of the clear and microbes free inhibitor zones begin within 24 hrs in bacteria culture plates and after 48-72 hrs in fungi. The antibacterial activity of each extract expressed in terms of diameter of zone of inhibition (mm) produced by respective extract (Upadhyay, 2016c) [38]. The findings were substantiated using advanced numerical tools and illustrated.

Table 1: Antibacterial and	Antifungal activities of	vegetable peel and	positive control antimicrobial	agents (Zone of inhibition in mm).

S. No.	Name of vegetable peel control positive	Antibacterial activities			Antifungal activities	
5. NO.	antimicrobial agents	E. coli	S. aureus	Proteus vulgaris	Lactobacillus	Saccharomyces cerevisiae
1.	Potato (Solanum tuberosum L.)	18	17	18	19	18
2.	Cucumber (Cucumis sativus L.)	22	21	19	20	14
3.	Sweet potato (Ipomoea batatas L.)	14	16	12	13	23
4.	Ginger (Zingiber officinale R.)	10	12	10	11	20
5.	Streptomycin (+ve control)	22	20	21	21	-
6.	Fluconazole (+ve control)	_	-	-	-	25

Results and Discussion

Antimicrobial activity was observed with different peels of vegetable against gram negative bacteria, gram positive bacteria and fungi. Zone of inhibitions marked the clear difference between the activities of different peels during the study (Table 1). As the bacteria and fungi belong to different groups and have structural and metabolic differences, therefore these vegetable peels differed in the activity accordingly. Amongst the selected fruits peels the highest antibacterial activity was found with cucumber against Escherichia coli (22 mm) followed by Staphylococcus aureus (21 mm), Lactobacillus (20 mm) and Proteus vulgaris (19 mm). The peak antifungal activity was recorded with sweet potato (23 mm) followed by ginger (20 mm), potato (18 mm) and cucumber (14 mm) against Saccharomyces cerevisiae in terms of zone of inhibition (Fig. 1).

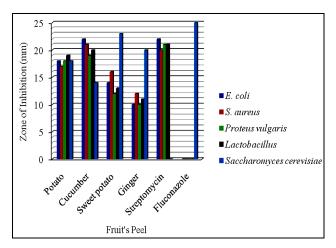


Fig 1: Antimicrobial activity (zone of inhibition in mm) of different fruits peel extracts against the selected microbes.

The differences in values of total phenolic compounds (TPC) were probably due to the color and variety of the potato tested (Al-Saikhan et al., 1995) [4]. A report on the presence of anthocyanins in the skin of colored potato varieties was found to be 2.5 fold higher than in the tuber pulp (Jansen and Flamme, 2006) [10]. Bracke et al., (1999) [6] reported that, flavonoids (naringenin, acycloguanosine, and tamoxifen compounds) from some plant extracts increased the antibacterial activity (Aggarwal et al., 2020; Yadav et al., 2020) [3, 39]. Some organic acids viz. citric, succinic, malic, acetic, and tartaric acids are commonly found in the fruit and vegetable peels having antimicrobial activity including Staphylococcus aureus (Prajapati et al., 2020) [15]. These attack cell walls, cell membranes, metabolic enzymes, protein synthesis systems, and the genetic material of microorganisms (Tripathi and Dubey, 2004) [33]. Numerous studies have provided evidences for decreased risk of some chronic diseases e.g., some types of cancer, cardiovascular and neurodegenerative disorders with increased dietary intake of vegetables, fruits, teas, spices and other plant-based foods and supplements (Sehrawat et al., 2020; Singh et al., 2020d; Tuli et al., 2021; Yadav et al., 2021) [20, 24, 35, 39]. The most abundant byproducts of minimal processing of fresh-cut fruit and vegetable were peel and seed that contain high amounts of phenolic compounds with antioxidant and antimicrobial properties (Shrikhande, 2000; Muthuswamy et al., 2008; Tuchila et al., 2008) [22, 14, 34]. Hence these studies confirmed that vegetable peel extracts consists of potential antimicrobial active compounds such as phenolic compounds, flavonoids and organic acids corroborated to the current findings. Thus these play a very significant role to control bacterial and fungal pathogens in ecofriendly and sustainable manner.

Conclusion

The vegetable peel is common household and industrial wastes which causes pollution problem if not utilized or disposed-off properly. This study is aimed to explore the new dimension applicability of vegetable peel wastes as potential source of low cost natural antimicrobial agent. The antimicrobial activities of potato, cucumber, sweet potato and ginger vegetable peel showed noticeable significant antimicrobial activities against the selected microbes. This investigation has open up the opportunity to these vegetable peels in drug development for the treatment and management of various pathogenic microbes. Therefore, the current findings will definitely unlock scope for utilization of the vegetable peel as sustainable and ecofriendly antimicrobial agents.

Acknowledgments

Authors are thankful to Prof. Anil K. Sharma, Head Dept. of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Mullana-Ambala (Haryana), India for kind permission and laboratory facility to perform the current research.

References

 Abdullah N, Kamarudin WSSW, Samicho Z, Aziman N, Zulkifli KS. Evaluation of *in vitro* antioxidant and antimicrobial activities of the various parts of *Benincasa hispida*. International Journal of PharmTech Research, 2012:4(4):1367-1376.

- 2. Abu Yazid N, Barrena R, Komilis D, Sánchez A. Solidstate fermentation as a novel paradigm for organic waste valorization: A review. Sustainability,2017:9(2):224.
- Aggarwal D, Upadhyay SK, Kaur L, Kumar A, Bhalla H, Singh R. Assessment of microbial burden on vegetable salads for food safety and human health. Bulletin of Pure and Applied Sciences-Zoology,2020:39A(1):130-136.
- 4. Al-Saikhan MS, Howard LR, Miller JC. Antioxidant activity and total phenolics in different genotypes of potato (*Solanum tuberosum* L). Journal of Food Science, 1995:60:341-343.
- 5. Bakkali F, Averbeck S, Averbeck D, Waomar M. Biological effects of essential oils- A review. Food Chemistry and Toxicology,2008:46(2):446-475.
- 6. Bracke, Depypere HT, Boterberg T, Van Marck VL, Vennekens KM, Vanluchene E *et al.* Influence of tangeretin on tamoxifen's therapeutic benefit in mammary cancer. Journal of National Cancer Institute,1999:91:354-359.
- 7. Burt S. Essential oils: Their antibacterial properties and potential applications in foods- A review. International Journal of Food Microbiology,2004:94:223-253.
- 8. Devi A, Dahiya VS, Upadhyay SK, Singh R, Sharma I, Kamboj P *et al.* Antimicrobial activity and phytochemical constituents in seed, leaf and bark extract of *Syzium cumini* (L). Plant Archives,2020:20(2):7787-7790.
- 9. Doharey V, Kumar M, Upadhyay SK, Singh R, Kumari B. Pharmacognostic, physicochemical and pharmaceutical paradigm of ash gourd, *Benincasa hispida* (Thunb.) fruit. Plant Archives,2021:21(1sp):249-252.
- Jansen G, Flamme W. Coloured potatoes (*Solanum tuberosum* L.)- Anthocyanin content and tuber quality. Genetic Resources and Crop Evolution, 2006:53:1321-1331.
- 11. Madhumithah CG, Krithiga R, Sundharam S, Changam SS, Guthakurta S, Cherian KM *et al.* Utilization of vegetables wastes for production of protease by solid state fermentation by using *Aspergillus niger*. World Journal of Agriculture Science, 2011:7(5):550-555.
- 12. Mahato N, Sharma K, Sinha M, Dhyani A, Pathak B, Jang H *et al.* Biotransformation of citrus waste- I: Production of biofuel and valuable compounds by fermentation. Processes, 2021:9:220.
- Massilia RM, Mosqueda-Melgar J, Soliva-Fortuny R, Martin-Belloso O. Control of pathogenic and spoilage microorganisms in fresh-cut fruits and fruit juices by traditional and alternative natural antimicrobials. Comprehensive Reviews in Food Science and Food Safety,2009:8:157-180.
- Muthuswamy S, Rupasinghe HPV, Stratton GW. Antimicrobial effect of cinnamon bark extract on Escherichia coli O157:H7, Listeria innocua and freshcut apple slices. Journal of Food Safety,2008:28:534-549
- 15. Prajapati PK, Aggarwal D, Upadhyay SK, Tuli HS, Yadav M, Sharma AK et al. Prevalence of inducible clindamycin resistance in methicillin resistant Staphylococcus aureus isolated from different clinical samples received in a tertiary care hospital. Bulletin of

- Pure and Applied Sciences-Zoology,2020:39A(2):344-353.
- Rajashekhara E, Tippannavar CM, Sreenivasa MN, Sharma JS. Inhibition activity of papain on facultative pathogens. Zentralblatt Fuer Microbiologie, 1990:145:455-456.
- 17. Rakholiya K, Kaneria M, Chanda S. Inhibition of microbial pathogens using fruit and vegetable peel extracts. International Journal of Food Sciences and Nutrition, 2014:65(6):733-739.
- 18. Saleem M, Saeed MT. Potential application of waste fruit peels (orange, yellow lemon and banana) as wide range natural antimicrobial agent. Journal of King Saud University-Science,2020:32(1):805-810.
- 19. Sathyabama S, Kinsley SJ, Sankaranarayanan S, Bama P. Antibacterial activity of different phytochemical extracts from the leaves of *T. procumbens*: Identification and mode of action of terpenoid compound as antibacterial. International Journal of Pharmaceutical Sciences, 2011:4:557-564.
- 20. Sehrawat N, Yadav M, Kumar S, Upadhyay SK, Singh M, Sharma AK *et al.* Review on health promoting biological activities of mungbean: A potent functional food of medicinal importance. Plant Archives, 2020:20(2 Spl):2969-2975.
- 21. Sheokand N, Sharma I, Singh R, Kamboj P. Antimicrobial activity and phytochemical analysis of *Catharanthus roseus* (L). Bio-Science Research Bulletin, 2019:35(2):53-57.
- 22. Shrikhande AJ. Wine byproducts with health benefits. Food Research International, 2000:33:469-474.
- 23. Singh A, Singh R, Navneet. Ethnomedicinal, pharmacological, antimicrobial potential and phytochemistry of *Trichosanthes anguina* Linn- A Review. Bulletin of Pure and Applied Sciences-Botany,2017:36B(2):82-90.
- 24. Singh C, Chauhan N, Upadhyay SK, Singh R. Phytochemistry and ethnopharmacological study of *Adiantum capillus-veneris* L. (Maidenhair fern). Plant Archives, 2020d: 20(2):3391-3398.
- 25. Singh M Renu, Kumar V, Upadhyay SK, Singh R, Yadav M. Biomimetic synthesis of silver nanoparticles from aqueous extract of *Saraca indica* and its profound antibacterial activity. Biointerface Research in Applied Chemistry,2021a:11(1):8110-8120.
- 26. Singh R, Upadhyay SK, Sunita. Phytodiversity of wild flora from Maharishi Markandeshwar (Deemed to be University), Mullana-Ambala, Haryana, India. Bulletin of Pure and Applied Sciences (Botany), 2018a:37B(2):130-136.
- 27. Singh R, Upadhyay SK, Kumar A, Rani A, Kumar P, Singh C *et al.* Lignin biodegradation in nature and significance. Vegetos,2018b:31(4):39-44.
- 28. Singh R, Upadhyay SK, Rani A, Kumar P, Kumar A, Sharma P *et al.* Ethanobotanical study of Subhartipuram, Meerut, Uttar Pradesh, India. I. Diversity and pharmacological significance of trees. International Journal of Pharmaceutical Research, 2019:11(4):782-794.
- 29. Singh R, Upadhyay SK, Tuli HS, Singh M, Kumar V, Yadav M *et al.* Ethnobotany and herbal medicine: Some local plants with anticancer activity. Bulletin of Pure and Applied Sciences -Botany,2020a:39B(1):57-64.

- Singh R, Upadhyay SK, Rani A, Kumar P, Kumar A. Ethanobotanical study of Subhartipuram, Meerut, Uttar Pradesh, India. II. Diversity and pharmacological significance of shrubs and climbers. International Journal of Pharmaceutical Research, 2020b:12(2):383-393.
- 31. Singh R, Upadhyay SK, Rani A, Kumar P, Sharma P, Sharma I *et al.* Ethnobotanical study of weed flora at district Ambala, Haryana, India: Comprehensive medicinal and pharmacological aspects of plant resources. International Journal of Pharmaceutical Research, 2020c:12(1SP):1941-1956.
- 32. Singh R, Upadhyay SK, Singh M, Sharma I, Sharma P, Kamboj P *et al.* Chitin, chitinases and chitin derivatives in biopharmaceutical, agricultural and environmental perspective. Biointerface Research in Applied Chemistry,2021b:11(3):9985-10005.
- 33. Tripathi P, Dubey NK. Exploitation of natural products as an alternative strategy to control postharvest fungal rotting of fruit and vegetables. Postharvest Biology and Technology, 2004:32:235-240.
- 34. Tuchila C, Jianu I, Rujescu CI, Butur M, Ahmadi-Khoie M, Negrea I *et al.* Evaluation of the antimicrobial activity of some plant extracts used as food additives. Journal of Food Agriculture and Environment, 2008:6:68-70.
- 35. Tuli HS, Mittal S, Aggarwal D, Parashar G, Parashar NC, Upadhyay SK *et al.* Path of silibinin from diet to medicine: A dietary polyphenolic flavonoid having potential anti-cancer therapeutic significance. Seminars in Cancer Biology, 2021. https://doi.org/10.1016/j.semcancer.2020.09.014.
- 36. Upadhyay SK. Anthelmintic and food supplementary conscientiousness of apitoxin in poultry model. Research Journal of Recent Sciences, 2016a:5(10):09-14
- 37. Upadhyay SK. Activity patterns of cell free supernatant of antagonistic microbial strains in rodents host-parasite systems. International Journal of Science and Research, 2016b:5(4):332-336.
- 38. Upadhyay SK. Allelopathic activities of specific microbial metabolites in the inland prawn fisheries off eastern Uttar Pradesh, India. International Journal of Scientific Research, 2016c:5(2):415-416.
- 39. Yadav M, Sehrawat N, Singh M, Upadhyay SK, Aggarwal D, Sharma AK *et al.* Cardioprotective and hepatoprotective potential of citrus flavonoid naringin: Current status and future perspectives for health benefits. Asian Journal of Biological and Life Sciences,2020:9(1):1-5.
- 40. Yadav M, Sehrawat N, Upadhyay SK, Kumar S. Emerging renoprotective role of citrus flavonoid naringin: Current pharmaceutical status and future perspectives. Current Pharmacology Reports, 2021:7(3):96-101.