



Evaluation of phytochemical constituents and antimicrobial activity of a novel polyherbal formulation

Fouzia Tehseen^{1*}, Syed Safiullah Ghori², Ruqiyah Khatoon¹, Qurrath Ul Ain Sana¹, Syed Mustafa Fawal¹

¹Department of Pharmaceutical Chemistry, Anwarul Uloom College of Pharmacy, Hyderabad, Telangana, India

²Department of Pharmacology, Anwarul Uloom College of Pharmacy, Hyderabad, Telangana, India

Abstract

The present study was performed for evaluation of antimicrobial activity of a novel polyherbal formulation (PHF) containing plants *viz.* Piper longum, Smilax china, Swertia chirata and Withania somnifera against bacteria (gram positive bacteria: S.aureus, B.subtilis and gram negative bacteria: E.coli and P.aeruginosa) and fungi (A.niger and C.albicans) responsible for various infections. The PHF was extracted using petroleum ether and ethanol to get three different concentrations of 25 µg/ml, 50 µg/ml and 100 µg/ml of each. Evaluation of phytochemical constituents revealed the presence of secondary metabolites like sugars, alkaloids, steroids, glycosides, tannins, saponins and flavonoids. Both the extracts were evaluated for antibacterial and antifungal activity by well diffusion assays. Ciprofloxacin and Miconazole were used as positive controls for antibacterial and antifungal activity respectively. All the extracts showed moderately significant antimicrobial activity. The 100 µg/ml ethanolic extract of the PHF was most effective against Candida albicans showing 30 mm zone of inhibition. The antimicrobial potency of the PHF may be due to the presence of tannins and polyphenols like flavonoids. The PHF showed synergistic broad spectrum antimicrobial activity providing scope for dosage formulation and further investigation for other pharmacological activities.

Keywords: polyherbal formulation, antimicrobial activity, extraction, phytochemical constituents, synergism

Introduction

Despite the advances in science for the treatment of diseases, man has been using plants as medicine. Plant based therapy has been used since ancient times and has attracted the attention of modern researchers to find the cure of various diseases. Traditional medicinal plants have received attention as the phytochemical constituents present maybe leads in the new drug discovery [1]. Herbal medication also called as botanical treatment or phyto medicine refers to the use of seeds, berries, roots, leaves, stem or flowers of a plant for medicinal purpose. Proper identification and appropriate quality with lack of adulteration, sophistication or substitution, is extremely important in the field of herbal medicine [2]. Researchers have been exploring plants as a potential source for the development of new leads as new therapeutic agents which are safe, efficient and less toxic. Ayurveda is one of the traditional medicinal systems with a history since ancient times. Ayurveda is known as Mother of all healing [3]. Allopathic medicines use mainly synthetic chemicals designed for specific target receptors which primarily give symptomatic relief whereas ayurveda uses natural means such as diet, herbs, minerals, spices, exercise, meditation, yoga, mental hygiene, sounds, smells and mechanical procedures to eliminate the root cause of the disease. It restores the balance and creates a healthy life style to prevent the reoccurrence of the imbalance. Ayurveda aims to integrate and balance the body, mind and spirit preventing illness, promoting wellness, longevity, vitality and happiness⁴. In Ayurveda, drug formulation can be as a single drug or use of more than one drug known as polyherbal formulation (PHF) [5, 6]. PHF exploits the strategy

of combining several medicinal herbs to obtain extra therapeutic effectiveness known as polypharmacy or polyherbalism [7]. PHFs are known to express high effectiveness in a number of diseases. The therapeutic effect of herbal medicines is due to the presence of a variety of phytoconstituents which is furthermore potentiated when compatible herbs are formulated together in PHFs. In a study in UK, the main reason for the usage of herbalism employing single herbs or PHFs is the effectiveness and favourable outcomes of the treatment [8]. Mostly PHFs have wide therapeutic range and are effective at low dose and safe at high dose. Antimicrobial agents are very important in reducing the global burden of infectious diseases [9]. Emergence of multidrug resistant strains in pathogenic bacteria have become a threat as there are fewer effective antimicrobial agents available for infection caused by pathogenic bacteria [10, 11]. A large number of medicinal plants are valuable resources of natural antimicrobial compounds which act as alternatively effective in treatment of resistant strains [12]. The literature revealed that Piper longum [13], Smilax china [14], Swertia chirata [15] and Withania somnifera [16] possess antibacterial and antifungal activity. The aim of the present work is to develop a novel polyherbal formulation which has potential antimicrobial activity using above four plants for a synergistic antimicrobial activity.

Material and Methods

Collection of plant material

The raw plant material for the PHF namely Piper longum, Smilax china, Swertia chirata and with aniasomnifera were bought from local market in Hyderabad and authenticated

by Dr. Abdul Samad, A.S Clinic, Hyderabad, Telangana, India. Nutrient agar and Sabouraud medium for antibacterial and antifungal activity were purchased from Standard chemicals, Hyderabad, Telangana.

Preparation of Polyherbal Formulation

Each of the plant was cleaned and made free of any earthy matter and powdered separately using a home grinder and mixed in proportions given in the table 1 and stored in an air tight glass container. The powdered PHF was extracted with petroleum ether and ethanol by hot extraction method. Soxhlet apparatus was used to carry out the hot extraction and the temperature was maintained between 60-80°C. Both the extracts were then dried at reduced pressure using rotavapour bath at 40-45°C. The dried extracts were dissolved in petroleum ether (PHFP) and ethanol (PHFE) to get 25 µg/ml (PHFP1), 50 µg/ml (PHFP2), 100 µg/ml (PHFP3) and 25 µg/ml (PHFE1), 50 µg/ml (PHFE2) and 100 µg/ml (PHFE3) which were prepared using distilled water. The above PHF extracts were stored in capped glass vials.

Evaluation of Phytochemical Constituents

Both the extracts were tested for the presence of phytoconstituents like carbohydrates, reducing sugars, alkaloids, sterols, flavonoids, saponins and tannins using standard qualitative tests [17].

Evaluation of Antimicrobial Activity

The antimicrobial activity was performed by agar well diffusion method. The bacterial strains employed were Gram positive bacteria- *Staphylococcus aureus* and *Bacillus subtilis*, Gram negative bacteria- *Escherichia coli* and *Pseudomonas aeruginosa*. The fungal strains employed were *Aspergillus niger* and *Candida albicans*. The microorganisms were obtained from culture collection of Microbiology Department, Anwarul Uloom college of Pharmacy.

Methodology of antibacterial activity [18]

To determine the antibacterial activity pure bacterial culture was sub cultured in nutrient broth at 37°C for 24 hours. About 100 µl of standardized inoculum of each bacterial strain was spread aseptically on the agar plates. With the help of a borer, cups/bore of about 6 mm diameter were aseptically made into the gelled agar plates. About 0.1 ml of different concentrations of the PHF extract are introduced into the wells/cups. Sterile DMSO was used as negative control. Ciprofloxacin (10µg/ml) was used as positive control. The plates were incubated for 24 hours at 37°C. The antibacterial activity was calculated by measuring the zone of inhibition. The zone of inhibition (ZOI) was measured to the nearest size in mm [19]. Appearance of ZOI is related to the presence of antimicrobial action in the PHF extracts.

Methodology of Antifungal activity [20]

To determine the antifungal activity of the different concentrations of PHF, pure fungi strains were sub cultured on Sabouraud's agar (SDA) at a temperature of 28°C for 3-5 days. A sterile borer was used to bore cups (6mm) in the SDA plates. Sterile DMSO was included as negative control while Miconazole (10µg/ml) was used as positive control. About 0.1 ml of the different concentration of PHF extract was transferred aseptically. The plates were incubated at 28°C for 2-3 days and then the ZOI was recorded and statistically analysed.

Results and Discussion

Results for Phytochemical evaluation of the PHF

The screening of extract revealed the presence of alkaloids, steroids, glycosides, tannins, saponins and flavonoids as shown in table 2.

Results for antimicrobial activity

The petroleum ether and ethanol extract of the polyherbal formulation exhibited moderate antimicrobial activity suppressing the growth of microbes as shown in table 3 and figure 1. The standard drugs exhibited higher antimicrobial activity against all test isolates when compared with both PHFP and PHFE. It was reported previously that ciprofloxacin is effective against *E.coli* and *P.aeruginosa*²¹. Evaluation of antimicrobial activity of the PHF extracts was recorded in table 3. Ethanolic extract of the PHF was found to have more potency when compared to the petroleum extract of the PHF. The PHFE3 extract showed moderately significant activity (21-30mm) against all the six strains of microbes followed by PHFP3 (17-26mm). Individual plants have various active phytochemical constituents or secondary metabolites which are responsible for the therapeutic activity exhibited by the plant²². The plants with varying potency when combined together as PHF may produce a greater potency as compared to the individual plant or sum of their individual effect known as the synergism²³. Tannins are known to inhibit many microbial enzymes in raw culture filtrates or in purified forms²⁴. The astringent property of the tannins is said to be reported due to its complexation with enzymes or substrates and metal ions²⁵. Polyphenols are said to have antimicrobial activity probably due to enzymes inhibition in the oxidised form or by more nonspecific interactions with the proteins. Flavonoids are hydroxylated polyphenolic compounds known to be produced by plants in response to microbial infections were studied extensively and are reported to show antimicrobial activity against many microorganisms²⁶. The PHF showed significant antimicrobial activity due to the presence of secondary metabolites like flavonoids and tannins that may be contributing to the antimicrobial action.

Table 1: Phytochemical Screening of Polyherbal Formulation

| S. No. | Phytoconstituent | Name of the test | Petroleum Ether extract (PHFP) | Ethanolic extract (PHFE) |
|--------|------------------|------------------------|--------------------------------|--------------------------|
| 1 | Carbohydrates | Mohlish test | - | - |
| 2 | Reducing Sugars | Benedicts test | - | + |
| 3 | Alkaloids | Dragendroffs test | + | + |
| 4 | Steroids | Salkowski test | - | + |
| 5 | Glycosides | Leibermanburchard test | + | - |
| 6 | Tannins | Keller killlani test | - | + |
| 7 | Saponins | Soap test | - | + |
| 8 | Flavonoids | Sulfuric acid test | + | + |

Table 2: Zone of inhibition of different concentration of PHF against microbial cultures.

| Concentration of the PHF extract (µg/ml) | Zone of Inhibition(mm) MEAN±SEM | | | | | |
|--|---------------------------------|-------------|--------------|---------------|-------------|-------------|
| | Test organism | | | | | |
| | B.subtilis | S.aureus | P.aeruginosa | E.coli | A.niger | C.albicans |
| Petroleum Ether | | | | | | |
| PHFP1 | 9.83±0.401 | 11.16±0.600 | 11±0.575 | 15.33±0.714 | 21±0.577 | 21.83±0.600 |
| PHFP2 | 11.66±0.421 | 13.5±0.763 | 13.16±0.703 | 18.5±0.428 | 22.83±0.600 | 22.83±0.307 |
| PHFP3 | 17±0.577 | 15.53±0.763 | 18.18±0.703 | 20.66±0.667 | 25±0.577 | 26.16±0.307 |
| Ethanol | | | | | | |
| PHFE1 | 10.83±0.307 | 11.5±0.763 | 11.16±0.6009 | 1.8336±0.401 | 22±0.577 | 22.5±0.428 |
| PHFE2 | 15.16±0.477 | 13.6±0.843 | 13.33±0.714 | 18.66±0.421 | 23.33±0.494 | 24.83±0.65 |
| PHFE3 | 21±0.461 | 18.66±0.843 | 18±0.575 | 24.66 ±0.494. | 29.5±0.562 | 30.33±0.557 |
| Standard(Positive Control) | | | | | | |
| Ciprofloxacin | 31.5±0.428 | 33.33±0.494 | 30.33±0.494 | 35.883±0.792 | - | - |
| Miconazole | - | - | - | - | 34.83±0.477 | 33.16±0.477 |

*Each value is Mean±SEM of 6 assays

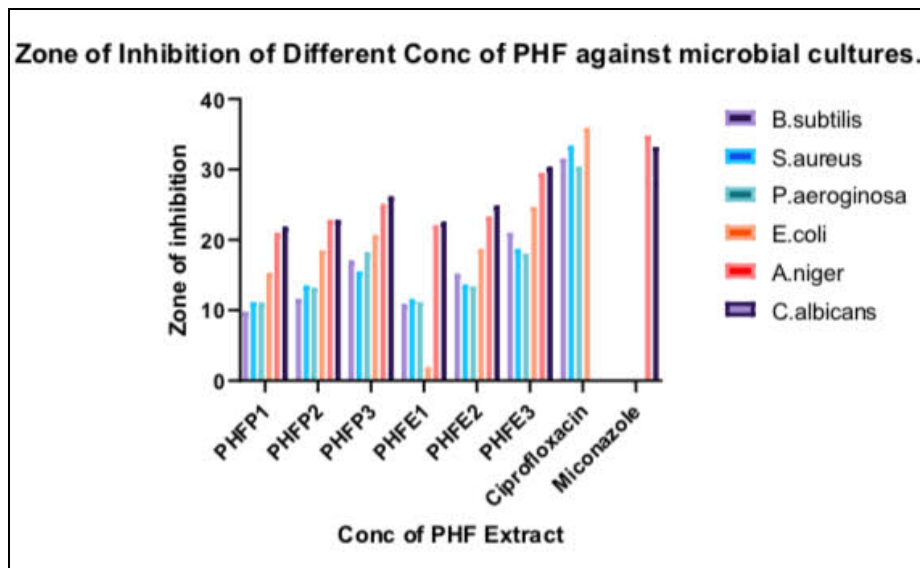


Fig 1: Antimicrobial activity of the Polyherbal Formulation.

Conclusion

The polyherbal formulation developed using a novel combination of plants viz. P.longum, S.china, S.chirata and W.somnifera demonstrated significant antimicrobial activity. Development of microbial resistance against antibiotics makes it essential need for effective medicinal plants as a cure from infectious diseases. Moreover PHF used in Ayurveda are plant products and are relatively cheaper, ecofriendly and readily available than the allopathic drugs. The study of herbal and medicinal plants as therapeutic agents is essential to explore the vast array of plant flora and know their efficacy in the treatment of diseases caused by microbes. Our research work can further be modified to get purified secondary metabolites and can be potential source for the discovery of new natural bioactive compounds.

References

1. Amor ILB, Boubaker J,Saghiar MB, *et al.* Phytochemistry and Biological activity of Plomis species. J. Ethanopharmacol,2009;125:183-202.
2. JE Robbers JE, Tyler VE. Tyler’s Herbs of choice: The therapeutic use of phytomedicinals. New York: Ed. Haworth Herbal. 1999, 287.
3. Kshirsagar M, Magno AC. Ayurveda – A quick reference handbook. USA: Lotus press, 2011.
4. Dahanukar SA, Thatte UM. Ayurveda revisited. Bombay: Popular Prakasan, 1989.
5. Jayakumar RV. Herbal medicine for type-2 diabetes. Int J Diabetes Dev Ctries,2010;30:111-112
6. Parasuraman S, Kumar EP, Kumar A, Emerson SF. Anti hyperlipidemic effect of triglize, polyherbal formulation. Int J Pharm Sci,2010;2:118-22.
7. Parasuraman S, Thing GS and Dhanaraj SA. Polyherbal formulation: Concept of Ayurveda. Pharmacogn Rev,2014;8(16):73-80.
8. Little CV. Simply because it works better: Exploring motives for the use of medical herbalism in contemporary U.K helath care. Complement Ther Med,2009;17:300-8.
9. Bhatia R and Narain JP. The growing challenge of antimicrobial resistance in the South- East Asia Region- are we losing the battle?. Indian Journal of Medical Research,2010;132(5):482-486.
10. Boucher HW, Talbot GH, Bradley JS *et al.* Bad bugs, no drugs: no ESKAPE! An update from the Infectious Disease Society of America. Clinical Infectious Diseases,2009;48(1):1-12.
11. Giamarellou H. Multidrug-resistnt Gram- negative bacteria: how to treat an for how long. International Journal of Antimicrobial Agent,2010;36(2):S50-S54.

12. Iwu MW, Duncan and Okunji CO. New antimicrobials of plant origin in perspectives on new crops and new uses. Plant breeding reviews Virginia, 1999.
13. Preeti Sreevstava. Therapeutic potential of *Piper longum* L. for disease management- A review. International Journal of Pharma Sciences,2014;4(4):692-696.
14. Shahrajabian MH, Sun W, Chen Q. Tremendous health benefits and clinical aspects of *Smilax china*. African Journal of Pharmacy and Pharmacology,2019;13(16):253-258.
15. Aleem A, Kabir H. Review on *Swertiachirata* as Traditional uses to its phytochemistry and pharmacological activity. Journal of Drug Delivery and Therapeutics,2018;8(5):73-78.
16. Singh G. Biological activities of *Withaniasomnifera*. Annals of Biological research, 2010;1(3):5-63.
17. Kokate CK, Gokhale SB. Practical Pharmacognosy, 2009.
18. Pelczar MJ, Chan ECS and Krieg NR. Microbiology: concepts and applications. McGraw publication., New York, 1993, 180.
19. National Committee for Clinical Laboratory Standard. Performance standards for Antimicrobial Susceptibility Testing Wayne PA,1999;9:M100-S9.
20. Fiori ACG, Schwan KRF, Stangarlin JR, Vida JB, Scapim CA, Guz MES, Pascholti SF. Antifungal activity of leaf extracts and essential oils of some medicinal plants against *Didymellabryonia*. Journal of Phtyopathology,2000;48:483.
21. Grillon A. Comparative activity of Ciprofloxacin, Levofloxaci and Moxifloxacin against *Klebsiella pneumonia*, *Pseudomonas aeruginosa* and *Stenotrophomonasmaltophilia* assessed by minimum inhibitory concentrations and time kill studies. PLOS ONE, 2016, 11(6).
22. Marjorie Mc. Lantproducers as antimicrobial agents. Microbiol Rev,1999;12:1053-1060.
23. Spinella M. The importance of Pharmacological synergy in psychoactive herbal medicines. Altern Med Rev,2002;7:130-7.
24. Jones GA, McAliister TA, Muir AD and Cheng KJ. Effects of sainfoin (*onobrychisvicifolia*) condensed tannins on growth and proteolysis of four strains of ruminal bacteria. App. Environ. Microbiol,1994;60:1374-1378.
25. Chung KY, Lu Z, Chou MW. Mechanism of inhibition of tannic acid and related compounds on the growth of intestinal bacterial. ChemRoxicol,1998;36:1053-1060.
26. Cowan MM. Plant products as antimicrobial agents. Clinical Microbiology reviews,1999;12(4):564-582.