



Anti-oxidative and hepatoprotective effects of methanolic extracts of seeds of *Brassica nigra* L., against CCL₄ induced hepatic damage in HEPG₂ cell lines

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Abstract

Accumulation of an excess amount of reactive oxygen species can cause hepatotoxicity that may result in liver damage. So development of antioxidant is needed for reducing liver toxicity. This study investigated the antioxidative and hepatoprotective activity of methanolic extract of seeds of *Brassica nigra* L., on carbon tetrachloride induced acute liver damage *in vitro*. The results of MTT assay indicated that methanolic extract of seeds of *Brassica nigra* L., was good antioxidant. The CCL₄ exposed HepG₂ cell line exhibited decreased cell viability, and elevated LPO, and decreased SOD, GSH and also increased the enzymes ALT, AST and LDH levels. Methanolic extract of seeds of *Brassica nigra* increased the cell viability and restoration of enzymes AST, ALT, LDH, and increased SOD, GSH levels and decreased the LPO and also restored the morphological changes of CCL₄insulted HepG₂celllines.

Keywords: *Brassica nigra* L., carbon tetrachloride, HepG₂cell lines, Liver enzyme markers

Introduction

Carbon tetrachloride is a industrial solvent that is used to induce chemical hepatitis and liver injuries in experimental animals. Carbon tetrachloride induced liver injuries are the most common experimental model for monitoring the hepatoprotective activity of certain drugs. A single exposure to CCL₄ as being a strong hepatotoxic xenobiotic directly leads to severe liver necrosis and steatosis^{1,2}. Mechanistic studies offer evidence that metabolism of CCL₄ via CYP2E1 to strongly reactive free radical metabolites plays a crucial role in the proposed mode of action. The major metabolites trichloromethyl (CCl₃[•]) and trichloromethyl peroxy ((CCl₃O₂)[•]) free radicals are extremely reactive and are capable of covalent bind to cellular macromolecules, preferring fatty acids of the membrane phospholipids. The free radicals induce cell membrane lipid peroxidation via disrupting polyunsaturated fatty acids within these membranes, initiating a sequential free radical chain reaction¹.

Materials and Methods

Cell culture³

HepG2 human liver hepatoma cells were obtained from National Centre for Cell Sciences (NCCS), Pune, India. Cells were grown in standard conditions: supplemented DMEM at 37°C in a humidified 5% carbon dioxide atmosphere. When the cells reached at 30,000 cells per well in a 96-well plates. 1x10⁶ cells per well in six plates, or 5x10⁷ cells per well in a single dish, depending on the determination. The cells were used after attachment.

CCL₄induced toxicity in HepG2 cells

HepG2 cells were incubated in medium or treated with toxic agent (30 mM CCL₄ in 0.05% DMSO). The evaluation assays were performed using standard methods.

Effect of Methanolic extract of *Brassica nigra* (L.), seeds on HepG2 cells

The hepatoprotective effects of Methanolic extract of *Brassica nigra* (L.), seeds on HepG2 cells were measured as follows; Normal control cells were incubated with DMEM in DMSO (0.05% v/v) for 12 h. For toxic treatment, cells were incubated with DMEM in DMSO (0.05% v/v) for 12 h and then treated with DMEM with 30mM CCL₄ for 1.5 h. For met-BN-S treatment cells were incubated with DMEM with met-S.BN at 150 µg/ml for 12 h and then treated with 30mM CCL₄ for 1.5 h

Cell viability assay

Cell viability was assessed using the MTT reduction assay with slight modifications. This colorimetric assay involves the conversion of MTT to purple formazan derivative by mitochondrial succinate dehydrogenase, which

is present only in viable cells. The cells were treated with met-BN-S and or the toxic agent. The medium was then removed and the cells were then incubated with MTT (0.5 mg/ml) for 2 h after which the formazan crystals were dissolved with 200 μ l/well of DMSO. Absorbance was measured at 570 nm. Viability was defined as the ration of the absorbance of treated cells to that of untreated control cells and is expressed as a percentage.

Measurement of AST, ALT and LDH activities

AST, ALT, and LDH activities were measured using an instrumentation Laboratory assay kits. HepG2 cells were treated with met-BN-S and or the toxic agent. The supernatant was removed from the wells. And the enzyme activities were measured immediately. The results are expressed as IU/L.

Measurement of GSH level

GSH level was quantified using a Glutathione assay kit. The assay kit is based on the enzymatic 5, 5'-dithiobis-2-(nitrobenzoic acid), disulfide dimer-oxidized GSH-reductase recycling method (DTNB). After treatment the medium was removed from the wells, and the adherent cells were scraped off and suspended in 0.5 ml of 50mM phosphate, pH 6.5, containing 1mM ethylene diamine tetra acetic acid sonicated and placed on ice. The supernatant of lysed cells was used to measure GSH level. Absorbance of the yellow product in the well was measured at a wavelength of 405 nm on a microplate reader at 5 min intervals for 30 min. the total GSH activity was measured using the kinetic method from a standard curve of GSH. The results are expressed as micromoles of GSH per litre.

Measurement of SOD activity

SOD activity was measured using a Superoxide Dismutase Assay kit, which uses a colorimetric assay to measure the concentration of formazan crystals. This assay uses a tetrazolium salt for the detection of superoxide radicals generated by xanthine oxidase and hypoxanthine. After treatment, the medium was removed from the wells, the adherent cells were scraped off and suspended in 20mM HEPES buffer, pH 7.2 containing 1mM EGTA, 210 mM mannitol, and 70mM sucrose, sonicated and placed on ice. To measure SOD activity the diluted radical detector and the supernatant of lysed cells or standard were added to each well of a 96 well plate, and xanthine oxidase was added. Absorbance in the well was measured at a wavelength of 460 nm after 20 min on a microplate reader. The results are expressed as IU/ml.

Measurement of Lipid peroxidation

The concentration of MDA the end product of lipid peroxidation was measured using a thiobarbituric acid reactive substance (TBARS) assay kit. After treatment the medium was removed from the wells, adherent cells were scraped off, suspended in cold PBS, sonicated and placed on ice. The supernatant from lysed cells or standard, sodium dodecyl sulphate, and the color reagent were added to each vial. The vial was heated at 100 C for 1 h and then immediately cooled in an ice bath and centrifuged; the content of each vial was transferred to a well in a microplate. The absorbance of the product was measured at a wavelength of 540 nm on a microplate reader. The extent of lipid peroxidation was quantified by estimating the MDA concentration. The results are expressed as micromoles of MDA equivalents formed per litre.

Measurement of IL-1 β & TNF- α (Pro-inflammatory cytokines) by ELISA

The levels of IL-1 β and TNF- α were assessed in cell lysates by using respective ELISA kits (Invitrogen, R&D systems & Alpha diagnostics, USA) as per the manufacturer' s protocol. Briefly, 150 μ l distilled water was added to the standard and blank wells for standard calibration and 100 μ l distilled water and 50 μ l of each supernatant were added in duplicate in to the wells. After incubation for 3 hour s at room temperature, the wells were emptied and washed three times with 150 μ l of wash buffer. TMB substrate (100 μ l) was added to each well and incubated for 15 min at room temperature followed by addition of 100 μ l stop- solution to all wells including blank wells. The absorbance was determined at 410nm using above mentioned ELISA reader (Roeske-Nielsen *et al.* 2004).

Statistical analysis

The data were analysed using one-way analysis of variance (ANOVA) Tukey's multiple comparison tests using Graph pad prism Version 6.0. Differences between means were considered significant at P< 0.05.

Results

Cell viability assessment – MTT assay [in presence of CCl₄ (30mM)]

Table 1: Effect of methanolic extract of seeds of *Brassica nigra* on cell viability level in CCl₄ intoxicated HepG2 cell lines.

S. No	Groups	% viability of cells
1	Control	99.13 \pm 0.210
2	CCl ₄ (30mM)	09.61 \pm 1.271***
3	Met-BN-S	61.14 \pm 3.287###

Statistical significance was determined by one way ANOVA followed by Tukey's multiple comparison tests using Graph pad prism Version 6.0. Values are represented as Mean \pm SEM, Superscript *** denotes $p < 0.001$ vs control, ### denotes $p < 0.001$; vs CCl₄ groups respectively, (CCl₄: Carbon tetrachloride).

Incubation of CCl₄ in HepG₂ cell lines have shown significant ($p < 0.001$) reduction in cell viability in comparison to control cells indicate the intensity of hepatotoxicity exhibited by CCl₄. Post-treatment with met-BN-S ($p < 0.001$) have shown notable increase in cell viability of HepG₂ cell lines evidence the protective effect exerted by extract in CCl₄ intoxicated condition.

Morphology

The control HepG₂ cells are tiny, highly light retractile, fibroblast-like and good morphological features and growing densely. CCl₄ (30mM) intoxication caused morphological changes with condensed cell density, swollen, decreased proliferation and differentiation of cells in comparison to the control cells. Post-treatment with Met-BN-S have significantly protected the cells from the CCl₄ toxicity which is evidenced by increased cell density and proliferation.

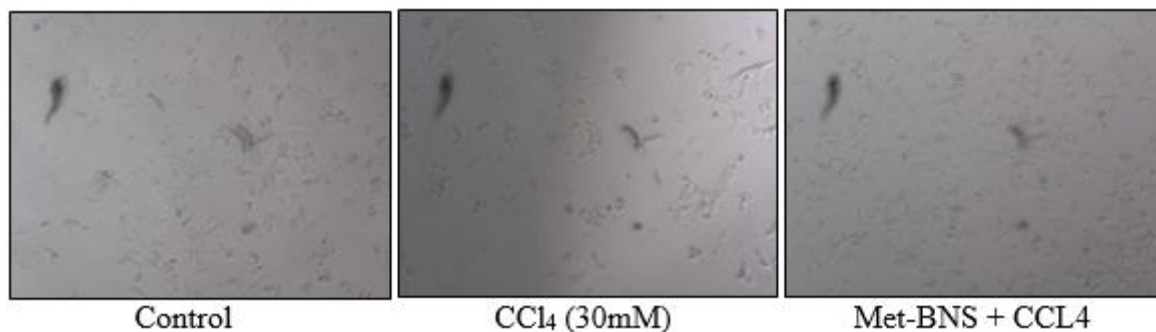


Fig 1

Table 2: Effect of methanolic extract of seeds of *Brassica nigra* (met-BN-S) on cellular LPO, GSH and SOD level in CCl₄ intoxicated HepG₂ cell lines.

S. No	Groups	LPO (n/mol)	GSH (n/mol)	SOD (n/mol)
1	Control	09.14 \pm 0.57	91.27 \pm 2.57	12.81 \pm 0.72
2	CCl ₄ (30mM)	76.89 \pm 1.27***	19.78 \pm 1.79***	1.27 \pm 0.96***
3	Met-BNS + CCl ₄	41.29 \pm 1.57###	65.67 \pm 3.57###	8.71 \pm 0.73##

Statistical significance was determined by one way ANOVA followed by Tukey's multiple comparison tests using Graph pad prism Version 6.0. Values are represented as Mean \pm SEM, Superscript *** denotes $p < 0.001$ vs control, ### denotes $p < 0.001$; vs CCl₄ groups respectively, (CCl₄: Carbon tetrachloride).

The SOD and GSH level ($p < 0.001$) was significantly reduced in CCl₄ intoxicated HepG₂ cell lines in comparison to control cells. Post-treatment with met-BN-S ($p < 0.001$) have remarkably elevated the SOD level in comparison to CCl₄ group indicate the free-radicals scavenging capacity of above extract in hepatotoxicity conditions. While the LPO level was significantly ($p < 0.001$) increased in CCl₄ intoxicated cell lines in comparison to control cells indicate the induction of oxidative stress. Post-treatment with met-BN-S ($p < 0.001$) have remarkably decreased the LPO level when compared to CCl₄ treated cells represent the reduced lipid peroxidation level in CCl₄ treated HepG₂ cell lines while treated with met-BN -S.

Table 3: Effect of methanolic extract of seeds of *Brassica nigra* (met-BN-S) on AST, ALT, LDH level in CCl₄ intoxicated HepG₂ cell lines.

Group	ALT(U/L)	AST(U/L)	LDH(U/L)
Control	8.53 \pm 2.55	12.51 \pm 0.45	290.25 \pm 0.62
CCl ₄ (30mM)	27.51 \pm 1.68***	34.55 \pm 1.10***	790.18 \pm 0.54***
Met-BN-S + CCl ₄	15.45 \pm 3.61###	24.35 \pm 1.31###	600.20 \pm 0.61###

Statistical significance was determined by one way ANOVA followed by Tukey's multiple comparison tests using Graph pad prism Version 6.0. Values are represented as Mean \pm SEM, Superscript *** denotes $p < 0.001$ vs control, ### denotes $p < 0.001$; vs CCl₄ groups respectively, (CCl₄: Carbon tetrachloride).

The LDH, ALT, AST level was significantly ($p < 0.001$) increased in CCl₄ intoxicated cell lines in comparison to control cells indicate the damage of hepatocytes that release LDH into culture medium. Post-treatment with met-BN-S ($p < 0.001$) have remarkably decreased the LDH, ALT, AST level when compared to CCl₄ treated cells represent the hepatoprotective effect of the met-BN-S on HepG₂ cell line

Anti-inflammatory assessments – ELISA**Table 4:** Effect of methanolic extract of seeds of *Brassica nigra* (met-BN-S) on IL-1 β and TNF- α level in CCl₄ intoxicated HepG₂ cell lines.

S.No	Groups	IL-1 β (pg/ml)	TNF- α (pg/ml)
1	Control	105.23 \pm 1.22	133.71 \pm 5.37
2	CCl ₄ (30mM)	889.87 \pm 17.89 ^{***}	977.84 \pm 36.52 ^{***}
5	Met-BN-S	623.87 \pm 32.65 ^{###}	675.24 \pm 18.14 ^{###}

Statistical significance was determined by one way ANOVA followed by Tukey's multiple comparison tests using Graph pad prism Version 6.0. Values are represented as Mean \pm SEM, Superscript *** denotes p<0.001 vs control, ### denotes p<0.001; vs CCl₄ groups respectively, (CCl₄: Carbon tetrachloride).

The IL-1 β , TNF- α level was significantly (p<0.001) increased in CCl₄ intoxicated HepG₂ cell lines in comparison to control cells indicate the inflammation of hepatocytes. Post-treatment with Met-BN-S have decreased the level of IL-1 β , and TNF- α level when compared to CCl₄ treated cells represent the reduced inflammatory response by above extract in CCl₄ treated HepG₂ cell lines.

Discussion

This research work deals with antioxidant and hepatoprotective activities of methanol extract of seeds of *Brassica nigra*. Liver is the second largest organ of our body involved in vital functions like detoxification of drugs and toxins through metabolism. CCl₄ is the toxin which produce trichloromethyl radical (CCl₃) which activated by cytochrome P450. This radical associated with CCl₄ induced hepatic damage. It reacts covalently with sulfhydryl-containing proteins in cells which leading to membrane lipid peroxidation and cell necrosis.

HepG₂ cell line was used to evaluate the hepatoprotective activity of methanolic extract of seeds of *Brassica nigra* (L.), against liver damage induced by CCl₄. Cells exposed to this toxic agents loss cell viability, release liver enzymes into the culture medium, do not metabolize the tetrazolium salt, and exhibit significantly changed levels of MDA, SOD and GSH⁷⁹⁻⁸⁰. We found that, compared with control cells, HepG₂ cells exposed to CCl₄ showed significantly reduced viability, reduced GSH and SOD levels and increased AST, ALT, LDH and MDA activities. Oxidative stress plays major role in CCl₄ toxicity. Oxidative stress occurs when the generation of reactive oxygen species overwhelms the ability to detoxify the reactive intermediates or exceeds the capacity to repair the resulting damage⁴. It has been reported that CCl₄ cause injury and reduce viability, increased AST, ALT, LDH. Enzyme levels are significant markers to determine the severity of damage to particular tissue. CCl₄ administration cause significant increase in enzyme levels like AST, ALT, LDH, which indicate the liver damage. Restoration of abnormal enzyme levels to normal indicating the protection of Methanolic extract of seeds of *Brassica nigra* against the liver injury by CCl₄.

It is important to mention that one way of indirectly assessing the damage to HepG₂ cells caused by free radicals is by measuring the activities of intracellular enzymes GSH and SOD and the viability of cultured cells by MTT assay. These measurements are useful for assessing the *in vitro* antioxidative actions of the hepatoprotective plant extracts^[5-9]. By-products of LPO cause hepatic damage in hydrophobic core of bio-membranes. Significant decline in LPO was observed in HepG₂ cell lines treated with methanolic extracts of seeds of *Brassica nigra*. We further studied non-enzymatic antioxidants like GSH and endogenous enzymes like SOD which involves in neutralizing free radicals. Suppression of these enzyme levels is an indicator for hepatic damage. CCl₄ administration cause decline in all the hepatic antioxidant levels in HepG₂ cell lines. Increased SOD and GSH levels was observed in HepG₂ cell lines treated with Met-BN-S indicates the antioxidant potential of the above extract which is a positive sign of liver protection. CCl₄ metabolism stimulated the Kupffer cells which activate TNF- α and IL-1 β is a strong inflammatory cytokines which involves in the production of prostaglandins^[10], macrophage activation and neutrophil infiltration. Activation of inflammatory cytokine cascade produces significant damage to the cells especially to hepatocytes. HepG₂ cell lines pre-treatment with Met-BN-S protects the hepatocytes from CCl₄ induced toxicity. Decreased TNF- α and IL-1 β in HepG₂ cell lines indicates the anti-inflammatory potential of the Met-BN-S.

Conclusion

We concluded that the methanolic extract of seeds of *Brassica nigra* offered significant hepatoprotection against the oxidative damage induced by CCl₄ in HepG₂ cells, as revealed by reduction of ALT, AST and LDH levels in the culture medium and remarkable decrease in intracellular LPO content, together with elevation of SOD activity and GSH concentration. Our recent study supported the *In vivo* study for the hepatoprotective effect of methanolic extract of seeds of *Brassica nigra*, thus further demonstrating that Met-BN-S could serve as a potent hepatoprotective agent in the food supplement, pharmaceutical and medical industries.

References

1. Manibusan MK, Odin M, Eastmond DA. "Postulated carbon tetrachloride mode of action: a review," Journal of Environmental Science and Health, Part C: Environmental Carcinogenesis and Ecotoxicology reviews, 2007;25(3):185-209.

2. Brautbar N, Williams J II, "Industrial solvents and liver toxicity: risk assessment, risk factors and mechanisms," International Journal of Hygiene and Environmental Health,2002:205(6):479-91.
3. Gonzalez *et al.*, *In vitro* assessment of hepatoprotective agents against damage induced by acetaminophen and CCl₄, Medicine, BMC Complementary and Alternative, 2017, 17-39.
4. Cover C, Mansouri A, Knight TR, Bjt ML, Lemasters JJ, Pessayre D, Jaeschke H. Per-oxynitrite induced mitochondrial and endonuclease-mediated nuclear DNA damage in acetaminophen hepatotoxicity. J Pharmacol Exp Ther,2005:315:879-87.
5. Torres-Gonzalez L, Munoz-Espibosa LE, Rivas-Estilla AM, Trujiilo-Murillo K, Salazar-sAranda R, Waksman De Torres N, Cordero-Perez P. Protective effect of four Mexican plants against CCl₄-induced damage on the Huh7 human hepatoma cell line. Ann Hepatol,2011:10:73-9.
6. Ghaffari H, Venkataramana M, Navaka SC, Ghassam BJ, Angaswamy N, Shekar S, Sampath Kumara KK, Prakash HS. Hepatoprotective action of Orthosiphon diffuses (Benth) methanol active fraction through antioxidant mechanisms; an *In vivo* and *In vitro* evaluation. J.Ethnopharmacol,2013:149:737-44
7. Pamplona S, Sa P, Lopes D, Costa E, Yamada E, E Silva C, *et al.* *In vitro* cytoprotective effects and antioxidant capacity of phenolic compounds from the leaves of Swietenia macrophylla. Molecules,2015:20(10):18777-88.
8. Liu JP, Feng L, Ahu MM, Wang RS, Zhang MH, Hu SY *et al.* The *in vitro* protective effects of curcumin and demethoxycurcumin in curcuma longa extract on advanced glycation end products-induced mesangial cell apoptosis and oxidative stress. Planta Med,2012:78(16):1757-60.
9. Mersch-Sundermann V, Knasmuller S, Wu XJ, Darroundi F, Kassie F. Use of a human –derived liver cell line for the detection of cytoprotective, antigenotoxic and cogenotoxic agents. Toxicology,2004:198:329-40.
10. Holden PR, James NH, Brooks AN, Roberts RA, Kimber I, Pennie WD. Identification of a possible association between carbon-tetrachloride-induced hepatotoxicity and interleukin -8-expression. J Biochem Mp; Toxicol,2000:14:283-90.