



Efficiency of Phosphate solubilizing bacteria isolated from different regions in dissolving of the insoluble phosphate and the activity of phosphatase enzyme

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Abstract

Ten isolates of Phosphate solubilizing bacteria were isolated from the soil of the rhizosphere for the plants cultivated in different locations of southern Iraq, the study of the culturing, microscopic and biochemical traits of these isolates, as well as testing the efficiency of these isolates in dissolving the phosphates in the liquid culture media, the activity of phosphatase enzyme, solubility index, their production of total acidity and reduce the reaction degree of the liquid media. The results showed that the isolates belonged to the *Bacillus* genus. B6 isolates were the most efficient isolates in dissolving the phosphate amounted to (36.6 mg P. L⁻¹) and solubility index amounted to (3.428), as well as their superiority in reducing the pH for the growth media from 7 to 5.6, their production to total acidity of (1.24 Meq.L⁻¹). The efficacy of the acid and alkaline phosphatase enzyme was (2.48 and 1.13 IU.L⁻¹), respectively.

Keywords: respectively, enzyme, production, phosphatase, superiority

1. Introduction

Phosphorus is considered the second most important mineral nutrient after nitrogen, which is needed by the plant and is specific for agricultural production because of its important role in the formation of cells and their division, the formation of cellular membranes, the transfer of genetic properties and other, where the Phosphorus availability is dependent on the change of the soil pH. In acid soils, phosphorus is deposited in it the form of insoluble iron phosphate and aluminum phosphate (Xaio, *et al.*, 2011) (Al-Taey *et al.*, 2017) [7]. Phosphorus is also less available in the alkaline media as a result of its deposition with calcium carbonate, thus most of the alkaline soil becomes containing on large deposits of accumulated phosphorus resulting from the successive annual additions of phosphate fertilizer (FNCA, 2006) [14]. In order to improve phosphorus availability in the soil, phosphorus solvents are tend to using the Phosphate solubilizing bacteria which have the ability to dissolve insoluble phosphorus compounds through acidizing, chelating and ion exchange (NIIR, 2007) [21]. It produces different types of organic and mineral acids and growth hormones such as Auxins, Gibberellins GA3 and Cytokinins, are considered the most important endogenous substances for modulating physiological and molecular responses, a critical requirement for plant survival as sessile organisms (Al-Taey, 2017, Al-Taey, 2018) [6-7, 5], Phytohormones act either at

their site of synthesis or elsewhere in plants following their transport thus reducing the harmful effects of nutrition imbalance resulting from environmental stresses (AL-Taey and AL-Musawi, 2019) [4]. As well as their ability to produce antibiotics that effectively contribute to plant protection from pathogens (Mohamed & Gomaa, 2012: Siddikee, *et al.*, 2011) [20, 26]. Phosphate solubilizing bacteria are considered one of the types of plant growth-promoting rhizobacteria (PGPR) which have different methods of promoting plant growth, such as fixing atmospheric

nitrogen, dissolving insoluble phosphorus, controlling some plant diseases, releasing some growth regulators, producing vitamins, enzymes, excretion of Osmosis resistance materials such as polysaccharides, amino acid, proline, acidic acid and ACC denminase, which improve the growth of growing plants under stress conditions (Omer, *et al.*, 2016) [22]. Inoculation with these bacteria reduces the use of mineral fertilizers and keeps the ecosystem from pollution (Vivekanandan *et al.*, 2015) [32]. Therefore, the study aims to isolate the Phosphate solubilizing bacteria from the rhizosphere soil samples of some crops and test their efficiency in dissolving the insoluble phosphates and the activity of phosphatase enzyme.

2. Materials and Methods

Rhizosphere soil samples for Alfalfa, tomato, wheat and barley plants were collected from the fields in Basra and Dhi Qar provinces. The plant was randomly taken off from each field at the flowering stage and obtained the soil of the Rhizosphere according to (Thaher, 2001) [30]. Phosphate solubilizing bacteria was isolated from different Rhizosphere soil samples by plate count dilution, using the modified solid Pikovskaya media (Sundara-Rao and Singh, 1963) [28]. After incubation of the plates at 28 °C for 2-3 days, the colonies of Phosphate solubilizing bacteria were identified through forming it a clear zone around the colonies as shown in Figure (1).

The solubility coefficient was estimated according method of (El-Komy, 2005) [13].

SI= D/C

Whereas

SI = solubility coefficient (without unit)

D = total diameter for the colony + transparent halo (cm)

C = diameter of the colony only (cm)

The culture traits of bacterial colonies were also taken and also microscopic and biochemical traits for isolates

(Bergey`s and Manual, 2004) [9].

The efficiency of the Phosphate solubilizing bacteria isolates was determined by using the free phosphate liquid Pikovskaya media, which was placed in 150 mL in a 250 mL flask and 1 g of tri-calcium phosphate was added as a source of insoluble phosphate. After sterilization, the flasks were inoculated with 1mL pure culture of Phosphate solubilizing bacteria, with age of 48 hr at a lighting density of 0.85 (Standard turbidity equivalent containing 1.5 x 10⁸ bacteria. ml⁻¹) according to (Baron and Finegold, 1990) [8], The flasks were incubated in the shaking incubator (100

cycles. min⁻¹) at 28 °C for 3 weeks and 10 ml was then taken from the liquid culture and filtered with centrifugal (6000 cycles / min). The pH for the media was estimated by the pH-meter as indicated by (Black, 1965a) [11] and the total acidity by the Titration was also estimated according to (Sberber, 1957) [27] by the following equation:

Percentage of total acidity =

$$\frac{\text{Normality of base} \times \text{volume} \times 0.064 \times \text{Final volume of the solution}}{\text{The volume of the Titrated solution} \times \text{Sample volume}} \times 100\%$$

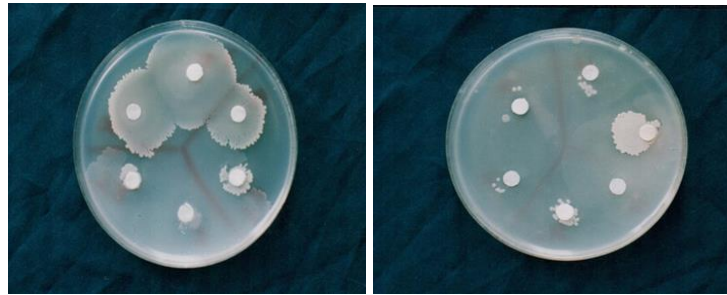


Fig 1: clear zone around colonies of Phosphate solubilizing bacteria isolated from rhizosphere soils for different plants

Dissolved phosphorus was estimated by spectrophotometer along a wavelength of 470 nanometers (Richards, 1954). The efficacy of acid and alkaline phosphatase was estimated by taking 0.5ml from the Pikovskaya media containing phosphate soluble isolates and 0.5 mL of (PNPP 0.115 M) P-nitrophenyl phosphate disodium substrate (Tabatabai & Bremmer, 1965) [29]. The activity of phosphatase was estimated according to the equation: (Al-Khan *et al.*, 2006) [3]

The activity of phosphatase enzyme = the intensity of light absorption for the model × 101

A 10 isolates of the Phosphate solubilizing bacteria were obtained for the rhizosphere samples of the roots obtained for the Alfalfa, tomato, wheat and barley plants cultivated in the soils of Basra, Dhi Qar, and Maysan provinces, which their cultivation, microscopic and biochemical traits listed in Table (1, 2), which showed that the isolates belong to the Bacillus genus (Holt *et al.*, 1984; Bergeys manul, 1984). Several studies have indicated that Iraqi soil is rich with Phosphate solubilizing bacteria isolates which belong to the Bacillus genus (Zahir, 1981; Hassan, 2013) [15].

Table 1: the cultivation and microscopic traits for Phosphate solubilizing bacteria isolates

Isolate code	Source of isolate	Cultivation description for bacterial isolates				Microscopic description for bacterial isolates			
		Color	The edge of the colony	Height of the colony	Size	Shape	Gram stain	movement	Aggregation
B1	Al-dawayya/ Alfalfa	Creamy	Regular	High	Middle	Short rod	+	motility	Pairs
B2	Al-bade / barley	White	irregular	High	Big	Short rod	+	motility	Chains
B3	Um Onaij/ Tomato	Creamy	Regular	High	Big	Short rod	+	motility	Chains
B4	Amarah / Alfalfa	White	Serrated	High	Big	Short rod	+	motility	Chains
B5	Al-dawayya/ Tomato	Yellow	Regular	High	Middle	Short rod	+	motility	Pairs
B6	Al-bade/ Alfalfa	White	Regular	High	Middle	Short rod	+	motility	Single
B7	Al Hartha/ Alfalfa	Creamy	Regular	High	Big	Short rod	+	motility	Chains
B8	Al-Zubair / Tomatoes	brouwn	irregular	High	Big	Short rod	+	motility	Chains
B9	Al-Rifai/ Alfalfa	White	Regular	High	Middle	Short rod	+	motility	Pairs
B10	Amarah / wheat	Creamy	Serrated	High	Big	Short rod	+	motility	Single

Table 2: Some biochemical traits for Phosphate solubilizing bacteria isolates

Isolate code	Catalase test	Oxidase test	Decomposition of starch	3% NaCl	37 ⁰	Methyl red	Nitrate reduction	Gelatin decomposition
B1	+	+	+	++	++	+	+	-
B2	+	+	+	++	+	+	+	-
B3	+	+	+	++	+	+	+	-
B4	+	+	+	+	+	+	+	-
B5	+	+	+	++	++	-	+	-
B6	+	+	+	++	++	-	+	-
B7	+	+	+	+	+	+	+	-
B8	+	+	+	+	+	+	+	-
B9	+	+	+	++	++	-	+	-
B10	+	+	+	++	++	+	+	-

Table 3: solubility Index of phosphate solubilizing isolates

Isolate code	Colonial diameter (cm)	Diameter of the transparent halo + colony (cm)	Phosphate Enzyme Index (SI)
B1	0.6	1.7	2.834
B2	1.0	2.1	2.100
B3	0.9	1.7	1.889
B4	0.9	1.8	2.000
B5	0.9	2.2	2.444
B6	0.7	2.4	3.428
B7	0.9	1.7	1.889
B8	0.9	1.6	1.778
B9	0.5	1.5	3.000
B10	0.8	1.8	2.250

Where all of the studied bacterial isolates varied in their ability to dissolve phosphate in the liquid and solid Pikovskaya. This is illustrated by the variation of the dissolving region diameters around the colonies as shown in Figure (1) and Table (3). The isolate B6 that isolated from the rhizosphere of the Alfalfa plant for the Al-bada region was the most efficient isolates in dissolving phosphate. The solubility index for it was 3.428, while the lowest average for dissolving phosphate was (1.778) by isolate B8 that

isolated from the tomato rhizosphere in Zubair region as shown in Table (3). This is due to the difference in the Phosphate solubilizing bacteria isolates in the quantity and quality of the metabolic compounds produced by organic acids such as Succinic acid, Glutamic acid, Malic acid, Fumaric acid and phosphatase enzyme activity, all of which work on dissolve phosphate compounds (Mardad *et al.*, 2013; Lin *et al.*, 2006)^[19, 18].

Table 4: Activity of alkaline and Acidic Phosphatase Enzyme (IU.L⁻¹) for Phosphate solubilizing isolates.

Isolate code	Acidic Phosphatase Enzyme activity (IU.L ⁻¹)	Alkaline Phosphatase Enzyme activity (IU.L ⁻¹)
B1	2.10	0.91
B2	1.87	0.59
B3	1.83	0.68
B4	1.80	0.77
B5	2.28	1.01
B6	2.48	1.13
B7	1.75	0.65
B8	1.69	0.73
B9	2.36	1.18
B10	2.00	0.18
Average	2.016	0.846

It is noted from Table (4) that Phosphate solubilizing isolate B6 showed significant efficiency in the activity of acid phosphatase enzyme by recording it (2.48 IU.L⁻¹), while isolate B8 was less effective for acid phosphatase enzyme was (1.69 IU.L⁻¹), whereas bacterial isolate B9 recorded significant excelling on the rest of the bacterial isolates in the activity of basal phosphatase enzyme amounted to (1.18 IU. L⁻¹), While B2 isolate recorded the lowest activity for basal phosphatase enzyme (0.59 IU. L⁻¹). These results agree with (Poumurugan and Gopi, 2006; Al-Jabouri 2016)^[23, 2] found that bacterial isolates differ in their efficacy to produce basal and acid phosphatase enzyme due to the genetic and environmental factors of the bacterial isolates themselves and found a positive correlation between the efficacy of the phosphatase enzyme and the amount of dissolved phosphorus. He *et al.*, (2004)^[16] indicated that Phosphate solubilizing bacteria possess phosphatase enzyme, which increases the concentration of metal phosphorus in the media from its low solubility sources, as well as that these bacteria conduct the process of organic phosphorus mineralization. The Phosphate solubilizing

bacteria isolate (B6) was excelled on all isolates in the reduction of pH to 5.60 during three weeks of incubation compared to the control treatment (non-inoculated) which gave pH 7.07 as shown in Table (5). This result agrees with (Alam *et al.*, 2004)^[1] that the low pH of the liquid media is due to the ability of isolates to produce organic acids such as citric acid, oxalic acid and 2-ketoGlucose acid, and that the quality and quantity of the acid produced is of great importance in dissolving the insoluble phosphate, These results agree with (Al-Jubouri, 2016; El-Komy, 2005)^[2, 13]. The bi-interaction between the different isolates and incubation period had a significant effect on the reduction of the degree of reaction for the liquid culture media, The highest decrease (5.22) was found in the interaction between isolate B6 and incubation period (21 days) while the lowest average (7.11) in the treatment of non-inoculation and incubation period 21 days. These results agree with (Salimpour *et al.*, 2010)^[25] that one of the most important indicators of Bacillus isolates activity is the low pH values of the culture media in which isolates grow over time.

Table 5: pH values in the liquid culture media inoculated with Phosphate solubilizing isolates treating with Tricalcium phosphate during various incubation periods

Isolate code	Incubation 3 days	Incubation 1 week	Incubation 2 weeks	Incubation 21 days	General average
Control	7.02	7.05	7.10	7.11	7.07
B1	6.56	6.38	5.82	5.89	6.17
B2	6.61	6.50	6.31	6.34	6.44
B3	6.70	6.62	6.053	6.55	6.60
B4	6.72	6.63	6.56	6.57	6.62
B5	6.10	5.097	5.56	5.53	5.79
B6	6.07	5.082	5.29	5.22	5.60
B7	6.75	6.071	6.64	6.62	6.68
B8	6.76	6.072	6.65	6.71	6.71
B9	6.12	5.091	5.50	5.43	5.74
B10	6.66	6.052	6.51	6.55	6.56
mean 6.56 6.44 6.23 6.23					
L.S.D _{0.05}	Isolate × Incubation period		Isolate		Incubation period
	0.824		0.412		0.275

The isolates also differed in their ability to produce total acidity in the liquid Pikovskaya media, where isolate B6 produced total acidity amounted to (1.33 meq.L⁻¹) after 21 days of incubation compared to the control treatment (non-inoculation) which amounted to (0.35 meq.L⁻¹) as shown in Table (6). This result agree with (Al-Jabouri, 2016) [2] increase in total acidity in the latter stages of incubation.

This is due to the use of Phosphate solubilizing bacteria in the metabolism, the construction of its cells and the formation of nucleic acids. It is also noted that the production of total acidity by bacterial isolates was highest during the first periods of incubation and began to decrease after two weeks of incubation and these results agree with (Zahir, 1981; Hassan (2013) [15].

Table 6: Total acidity values in the liquid culture media inoculated with Phosphate solubilizing isolates treating with Tricalcium phosphate during various incubation periods

Isolate code	Incubation 3 days	Incubation 1 week	Incubation 2 weeks	Incubation 21 days	General average
Control	0.47	0.43	0.39	0.35	0.41
B1	0.88	1.02	1.10	1.04	1.01
B2	0.78	0.88	0.93	0.89	0.87
B3	0.73	0.83	0.86	0.82	0.81
B4	0.72	0.85	0.86	0.85	0.82
B5	0.97	1.20	1.28	1.27	1.18
B6	1.010	1.22	1.31	1.33	1.24
B7	0.70	0.78	0.80	0.80	0.77
B8	0.67	0.74	0.78	0.77	0.74
B9	1.02	1.19	1.29	1.30	1.20
B10	0.79	0.86	0.90	0.85	0.85
mean 0.80 0.91 0.96 0.93					
L.S.D _{0.05}	Isolate × Incubation period		Isolate		Incubation period
	0.216		0.108		0.072

Bacterial isolates differed significantly in the dissolve of tricalcium phosphate in the liquid culture media as shown in Table (7). The B6 isolate was excelled in their solubility for phosphate on the other isolates amounted to (38.7 mg P.L⁻¹) after three weeks of incubation, while the lowest average was (4.8 mg P.L⁻¹) when untreated at the same time period.

This is due to the ability of these isolates to secrete the various organic acids that dissolve the insoluble phosphate (EI-Komy, 2005) [13]. The greatest effect is due to the type of acid produced by bacteria such as lactic acid (Didiek *et al.*, 2000) [12].

Table 7: The amount of dissolved phosphorus (mg P.L⁻¹) in the liquid culture media inoculated with Phosphate solubilizing isolates treating with Tricalcium phosphate during various incubation periods

Isolate code	Incubation 3 days	Incubation 1 week	Incubation 2 weeks	Incubation 21 days	General average
Control	6.2	5.8	5.1	4.8	5.48
B1	21.3	25.1	31.6	29.0	26.75
B2	17.1	21.0	22.7	21.2	20.50
B3	11.3	18.2	19.0	17.5	16.50
B4	11.1	19.0	19.2	17.7	16.75
B5	22.3	34.0	37.1	35.6	32.25
B6	32.0	35.3	38.0	38.7	36.00
B7	10.1	17.0	18.2	16.7	15.50
B8	11.2	13.1	17.7	17.0	14.75

B9	25.4	34.3	38.0	37.3	33.75
B10	17.1	18.6	21.1	18.2	18.75
Mean 16.83 21.95 24.34 23.06					
L.S.D _{0.05}	Isolate × Incubation period		Isolate		Incubation period
	7.652		3.826		2.381

References

- Alam MS, Cui ZJ, Yamagishi T, Ishir R. Grain yield and related physiological characteristics of rice plants (*Oryza sativa* L.) inoculated with free living rhizobacteria. *Plant, Prod. Sci.* vol. 2004; 4(2):126-130.
- Al-Jubouri, Abdullah Kareem Jabbar. Isolate of *Pseudomonas fluorescens* and *Bacillus subtilis* bacteria and their diagnosis by PCR technique and study their effect on the pollen technique stabilized in the growth and yield of yellow corn (*Zea mays* L.). PhD thesis, College of Agriculture, University of Baghdad, 2016.
- Al-khan Hussein Ismail, Haitham L Al-Hayali, Thaer M Al -Mushhadani. Effect of Aqueous Extract of Nerium oleander and Melia azedarach Plants on the Morphology and Osmolarity of *Leishmania tropica* Promastigotes *in Vitro*. *Journal of Science Rafidain.* 2006; 17(11):68-79.
- Al-Taey DKA, Almusawi ZJM. Effect of nano-fertilizers, salicylic acid, and organic matter in growth and yield of rocket (*Eruca sativa* Mill) under Salt stress. *International Journal of Botany Studies.* 2019; 4(3):77-81.
- Al-Taey DKA, Majid ZZ. Study Effect of Kinetin, Bio-fertilizers and Organic Matter Application in Lettuce under Salt Stress. *Journal of Global Pharma Technology.* 2018; 10(1):148-164.
- Al-Taey DKA. Mitigation of Salt Stress by Organic Matter and GA3 on Growth and Peroxidase Activity in Pepper (*Capsicum annum* L.). *Advances in Natural and Applied Sciences.* 2017; 11(10):1-11.
- AL-Taey DKA, Al-Janabi ASH, Rachid AM. Effect of water salinity, Organic and minerals fertilization on growth and some nutrients elements in cabbage *Brassica oleracea* var capitata. *Babylon journal of Pure and Applied science.* 2017; 25(6):232-248.
- Baron EJ, Finegold SM. *Diagnostic Microbiology.* 8th ed. C. V. Mosby Company. USA, 1990.
- Bergey Manual S. *Systematic Bacteriology.* Williams and Wilking. Baltimore. London, 2004.
- Bergey DH, Buchanan RE, Gibbson NE. *Manual of determinative bacteriology.* 8th Edition, Baltimore: the William and Wilkins Company, 1974.
- Black CA. *Methods of soil Analysis part (1). Physical Properties* Am. Soc. Agron. INC. Publisher, Madison, Wisconsin, U.S.A, 1965a.
- Didiek Siswanto HG, Sugiarto Y. Bioactivation of Poorly Soluble Phosphate Rocks with a Phosphorus Solubilizing Fungus. *Soil Sci. Soc. of Amer. J.* 2000; 64:927-932.
- El-Komy HM. Coimmobilization of *Azospirillum lipoferum* and *Bacillus megaterium* for successful phosphorus and Nitrogen nutrition of wheat plants. *J. Food. Technol. Biotechnol.* 2005; 43(1):19-27.
- FNCA. *Forum for Nuclear Cooperation in Asia, Biofertilizer mamud,* 2006.
- Hassan Zeinab Kadhim. Isolate and diagnosis of *Azospirillum lipoferum* and *Bacillus polymyxa* bacteria from some soil in southern Iraq and their role in the bio-fertilization for yellow corn plants (*Zea mays* L.). PhD thesis. College of Agriculture. Basra University, 2013.
- He ZQ, Griffin TS, Honeycutt CW. Enzymatic hydrolysis of organic phosphorus in swine manure and soil. *J. Environ. Qual.* 2004; 33:367-372.
- Holt JG, Kreig NR, Sneath PH, Staley JT, Williams ST. *Bergey's Manual for Determinative Bacteriology.* 9th ed. Williams and Wilkins U.S.A. P. 1994; 93:151-155.
- Lin TF, Huang HI, Shen FT, Young CC. The protons of gluconic acid are the major factor responsible for the dissolution of tricalcium phosphate by *Burkholderia cepacia* CC-A174. *Bioresour. Technol.* 2006; 97:957-960.
- Mardad Ilham Mardad, Aurelio Serrano, Abdelaziz Soukri. Solubilization of inorganic phosphate and production of organic acids by bacteria isolated from a Moroccan mineral phosphate deposit. *African Journal of Microbiology Research.* 2013; 7(8):626-635.
- Mohamed HI, Gomaa EZ. Effect of plant growth promoting *Bacillus subtilis* and *Pseudomonas fluorescens* on growth and pigment composition of radish plants (*Raphanus sativus*) under NaCl stress. *Photosynthetica.* 2012; 50(2):263-272.
- NIIR. *The complete technology book on bio-fertilizer and organic farming,* 2007.
- Omer AM, Emara HM, Zaghloul RA, Monem MOA, Dawwam GE. Potential of *Azotobacter salinestris* as plant growth promoting rhizobacteria under saline stress conditions. *Research Journal of Pharmaceutical, Biological and Chemical Sciences.* 2016; 7(6):2572-2583.
- Ponmurugan P, Gopi C. *In vitro* production of growth regulators of phosphatase activity by phosphate solubilizing bacteria. *Afr. J. Biotechnol.* 2006; 5:348-350.
- Richards LA. *Dignosis and Improvement of saline and Alkali soils.* U.S. Dept. Agr. H.B. No, 1954, 60.
- Salimpour Khavazi SK, Nadian H, Besharati H, Miransari M. Enhancing phosphorous availability to canola (*Brassica napus* L.) using P solubilizing and sulfur oxidizing bacteria. *Australian Journal of Crop and science.* *AJCS.* 2010; 4(5):330-334.
- Siddikee MA, Chauhan PS, Anandham R, Gwang-Hyun Han, Tongmin sa. Isolation, Characterization, and Use for Plant Growth Promotion under Salt Stress, of ACC Deaminase-Producing Halotolerant Bacteria Derived from Coastal Soil. *J. Microbiol. Biotechnol.* 2011, 1577-1584.
- Sperber JI. *Nutr. (Cited by Introduction of soil Microbiology, 1982-1957; 180:994-995.*
- Sundara-Rao Sinha JP. Solubilization of phosphate by phosphorous solubilizing organisms using P32. At trace and influence of seed bacterization on uptake by crop. *J. Indian. Soc. Soil Sci.* 1963; 11:209-219.
- Tabatabai MA, Bremmer JM. Use of P. nitrophenyl phosphate for assay of soil phosphatase activity. *Soil boil. Biochem.* 1965; 1:301-307.

30. Thaher Abdal Zahra Taha. Response of yellow corn plants (*Zea mays* L.) to vaccinate some locally isolated *Azospirillum brasilense* bacteria. PhD thesis, College of Agriculture, University of Baghdad, 2001.
31. Thaher Abdal-Zahra Taha. Study of phosphate soluble microorganisms in the root zone of some crops. Master Thesis, University of Baghdad, 1981.
32. Vivekanandan M, Karthik R, Leela A. Improvement of crop productivity in saline soil through application of saline- tolerant rhizosphere bacteria – Current Prospective. Inter. Journal of Advanced Research V. 2015; 3(7):1273-1283.
33. Xiao CQ, Chi RA, He H, Qiu GZ, Wang DZ, Zhang WX. Isolation of phosphate solubilizing fungi from phosphate mines and their effect on wheat seedling growth. Appl. Biochem. Biotechnol. 2011; 159:330-342.